

Table 2c. Potential risks and management options for clonal banks.

Activity	Risk Sources/Indicators	Risk/Consequence	Action Plan		
			People	Facility	
ACQUISITION					
Collecting	Narrow genetic variability and large gaps in germplasm collection	Failure to capture diversity in field	Send Center genebank personnel to take the lead in joint collecting missions with national programs. Conduct training of collectors in partner countries. Maintain and hire a pool of expert collectors.		Analyse collector conduct gap-filling
	Untrained personnel in collecting and documentation	Failure to capture diversity in field and document important information	Send Center genebank personnel to take the lead in joint collecting missions with national programs. Conduct training of collectors in partner countries. Maintain and hire a pool of expert collectors.		
	Misidentification of germplasm	Misleading information	Include taxonomists during collecting.		
	Lack of simple collection protocol and documentation forms	Failure to capture diversity in field			Develop simple c
	Agricultural intensification, replacement of traditional varieties with modern ones, urbanization, land use change, and climatic events	Loss of germplasm in habitat			Prioritize affected that can fill gaps i
	Strict country and international laws on access and use of germplasm	Poor access and use of germplasm in unexplored areas			Secure a Germpl between donor cc accessions under
	Breach of country and international treaties	Legal consequences. Damaged reputation and relationship	Training of all institute staff on internationally agreed protocols, in consultation with Genebank and other Center authorities.		Follow national pr permits, under rel Collect in partners
	Ambiguous position of countries regarding international treaties	Poor access and use of germplasm in unexplored areas			Foster goodwill th breeding and Tre and incentivize dc
Donation	Received foreign materials carry pests and diseases	Introduction of pest and diseases to host country			Strictly observe qu main storage area decontaminated. (screenhouse or a
	Limited germplasm testing capability	Restricts international germplasm exchange		Develop testing and handling capability for pests and diseases of international importance.	
	Reluctance to share germplasm due to IP rights	Restricts international germplasm exchange		Conduct training on benefits and limitations of IP rights.	
	Working collections not duplicated in major genebanks	Failure to capture elite germplasm			Proactively conse
CONSERVATION					
Registration	Unverified passport and other data submitted	Incorrect or unreliable passport data, and poor quality of scientific reports			Verify passport in
	Received materials have low viability	Loss of germplasm			Obtain large amo properly.

	Limited storage space for clonal materials				Priority materials : a) cultivars and elite clones b) clones from certain diploid forms) c) clones from sex selection d) clones with unique resistances e) highly diverse clones with molecular markers Conduct regular re-evaluation of materials
Conservation in <i>in vitro</i> Banks					
<u>Sample Processing</u>	Untrained or inefficient personnel in sample processing	Reduction of good quality propagules and accidental mixtures	Conduct regular training and enforce close supervision of personnel on detection and removal of infected, infested and mechanically damaged samples.		Subculture samples (separated in time) to spare cultures of fresh new subcultures and
	Source of material is infected	Loss of viability of propagules		Provide an isolated growth room for <i>in vitro</i> explants taken directly from the field to allow time to detect insect infestations and disease infection and prevent their spread to other cultures.	Use material from field records database records to ensure procedures can be followed. Use surface disinfection (0.5 to 1% or more) on explants in a fungicide solution. Additional treatment with sodium hypochlorite or dips, and insecticide before explants are wrapped during transport.
	Poor quality and/or suboptimal size of propagule	Loss of meristems			Have additional media and growth conditions for experimentation to test for moderate growth.
	Weak mother plants	Short lifespan of propagules in storage			Collect plant material from vigorous and healthy mother plants for <i>in vitro</i> introduction.
	Ineffective pest and disease screening procedures during sample processing	Reduction of good quality propagules			If an isolation area is used, ensure that it is free from the field shortly after mites and thrips from the field.
	No efficient tissue sterilisation procedures	Poor quality of propagules			Conduct research on tissue sterilisation procedures.
	Lack of proper disposal procedures of contaminated plant materials	Increase in <i>in vitro</i> contamination with pests and diseases and dissemination to new areas			Autoclave all contaminated materials before discarding or cleaning areas.
	Ineffective thermotherapy procedure	Failure of explants to multiply			Conduct experiments on thermotherapy procedures for moderate growth. Treat materials sequentially. The first group is damaged by the first treatment.
	Inappropriate media and conditions for culture initiation	Failure of explants to multiply			Conduct experiments on media composition and culture conditions for growth and multiplication. Use a triage system and
<u>Germplasm Testing</u>	Untrained personnel in health testing of propagules	Pest and disease damage and spread in collection	Train staff to be observant of unusual growth or symptoms in the cultures.		

	Improper screening methods and monitoring regime	Pest and disease damage and spread in collection		Conduct regular monitoring of the cultures, storage rooms and growth room. Use a pyrethrum-based spray in culture rooms. Regularly check all sterilisation equipment and laminar air flow quality	Have a monitoring sources. Conduct on a regular basis and before storage treatments. Remove cultures, unless they are of the germplasm potentially contaminated to minimize spread
	Microbes and pests are not apparent at initial testing but appear later.	Pest and disease damage and spread in collection			Test at explant intervals.
	Untrained personnel in transgene detection	Loss of genetic integrity of other accessions			
	Inadvertent presence of transgene	Loss of genetic integrity of other accessions			
	Lack or improper determination of transgene presence	Inaccurate or wrong information regarding transgene presence			
	Limited quantity of high quality propagules	Loss of accession			Monitor plants in 3 months to assess necrosis, chlorosis, callus formation and of viable cultures. If a certain percentage decreased, subset of fresh tissue process, monitor ;
	Ineffective sterilization techniques	Loss of accession			To decrease contamination with 70% ethanol or prior soapy water sterilizers instead
	Misapplication of antibiotics	Loss of accession			Apply short (10 days) antibiotics in the culture to control contamination. Add media to control fr
	Somaclonal variation	Loss of genetic integrity			Use appropriate number of in vitro collected somaclonal variations for maturity in field or morphological characteristics. Monitor somaclonal variations closely. Develop variation detection as reference samples
Conservation Procedure	Errors in media preparation	Loss of accession			Use specific protocols and document down all steps to ensure necessary.
	Ineffective pre-treatment	Short lifespan of propagules in storage			Apply two weeks of cold in the normal growth cold acclimatization
	Chemical imbalance during culture	Abnormal growth of material			Check cultures for necrosis and browning in the field if needed. Ensure proper balance of cytokinins during propagation, which
	Suboptimal culture methods for a broad range of genotypes				Conduct additional tests and techniques suitable

	Short storage life of propagules	Loss of viability			Develop methods collections. Adjust samples to extent where there is a r
	Delayed inventory	Loss of material			Schedule inventory between which re genus.
	Late subculturing	Loss of viability			Conduct regular n culturing as when been reduced to 3
	Backlog in regeneration	Loss of viability			Periodically check performance of st recommended int pot plants in the g experienced.
<u>Storage Facility</u>	Unsterile transfer facilities	Loss of accession		Design transfer facilities with minimal foot traffic and outside airflow.	Regularly check c equipment (smok plates. Check for hoods are moved
	Unsuitable tissue culture containers for in vitro samples	Loss of accession			Carefully seal indi vitro samples wi contamination, pe replicates in sepa container-specific
	Poor laboratory maintenance	Contamination and loss of materials	Field and greenhouse personnel should change their shoes and clothing before entering the lab and growth rooms.	Routinely mop floors with disinfectant. Control dust and insects, especially mites. Regularly change or clean filters in the laminar flow hoods and building's ventilation system.	Autoclave contam washed or remov washing area. Ne separate room or plastic wrap until t out. Wipe cultures laboratories with i isolated from the i insect infestations
<u>Safety Duplication</u>	Safety duplication site is vulnerable to natural calamities	Inaccessible or loss of safety duplication			Store duplicates in either on-site in sr box or active colle duplicate as base (cryo).
<u>Regeneration</u>	Regeneration failure	Loss of germplasm			Adhere strictly to procedure.
Conservation in Cryo bank - Long Term Storage (LTS)					
<u>Sample Processing</u>	Incorrectly identified material is stored	Wrong germplasm stored and distributed			Use only verified
	Isolation of material is not done correctly, meristems are damaged and regrowth as callus	Increased chance of variation	Training of lab personnel		
	Chemical cryoprotectants injure plant cells during pre-treatment	Reduced viability during storage			Optimize procedu
	Plants are sensitive to preculture method	Loss of viability			Choose another p
	Technique does not work for all plants in the collection	Gaps in collection			Plan to have sever
<u>Germplasm Testing</u>	Thawing/rewarming is done improperly	Underestimate of post-thaw regeneration rate	Training for staff		Have standard pr
	Water bath may be contaminated	Damage to samples			Use sterile water waterbath.
	New material in cryo-collection is not viable	Loss of samples			Conduct viability t and have a written regrowth (mediurr
<u>Conservation Procedure</u>	Dewars may fail.	Damage to samples			Use alarm system separate dewar.

	Unreliable supply of liquid nitrogen (LN)	Damage to samples			Ensure a reliable companies, local insemination cent manufacturing pla hold dewars and p supply.
	Rapid loss of LN in dewar	Damage to samples			Provide a wide-m during processing dewar fo long-term Liq N, with autom when limits are re and fill dewars req
	Improper placement on cryocane and to multiple rewarming and cooling cycles during sample retrievals	Loss of biological stability			Follow instruction dewar. Group san Make more replic demand. Store lo often used sampl
	Compromised integrity of cryovials	Contamination and loss of biological stability			Use cryovials with as cap-threads, a cryosleeves, and
	Insufficient number of stored propagules	Loss of germplasm			Determine the nu based on the surv speed of propaga storage.
Conservation on field banks					
Sample Processing	Low initial quality of explants.	Short lifespan of germplasm in storage			Collect plant mate and healthy moth
	Improper conditioning and propagation of vegetative material	Short lifespan of germplasm in storage			Conduct immedia disinfection, depe
	Failure in propagation and storage of propagules	Loss of germplasm			Group accessions procedures. Carr do not respond w Contact other faci information on pro
Germplasm Testing					
<i>Health Diagnosis</i>	Failure to detect and remove samples with pests and diseases and improper disposal of contaminated materials	Increased pathogen or pest population in the facility, thereby jeopardizing the health of other accessions in the collection as well as introducing new pest or diseases in new regions/countries.	Conduct regular training and enforce close supervision of personnel on proper disposal of contaminated materials.	If applicable, grow incoming and regeneration materials in screenhouse or in isolation away from large areas of local farms. Duplicate collection in two other sites, or keep an in vitro or a cryo set.	Subject regenerat phytosanitary test established, incin sterilize and disca regularly and imm Accessions with s diseases or pests such as being pla being treated for t schedule.
	Ineffective screenhouse to control insects			Construct and manage screenhouses to prevent disease-carrying insects from entering. Workers and visitors should not enter the screenhouses after visiting field plots. The entryway into the screenhouses should have a set of two doors that should not be opened at the same time to reduce the entry of insects. Check screens and structures periodically to assure they remain insect proof.	
	Backlogs in pest and disease monitoring	Loss of field bank samples	Hire and train adequate personnel to regularly monitor pest and diseases.		
	False positive and false negative results during plant health testing.	Loss of materials due to false positive results. Dissemination of diseased materials due to false negative results.			Repeat tests in ca to confirm and ha

<i>Storage Monitoring</i>	Limited numbers of viable plants	Loss of germplasm			Keep 3 to 20 veg
	Mechanical mixtures or invasive plants	Loss of genetic integrity			Monitor the plants immediately.
	Late rejuvenation or multiplication (plants lost their physiologic vigour or accumulated pests and diseases)	Loss of materials			Monitor the genet regeneration in ac
<i>Conservation Procedure</i>	Inadequate selection, pre-conservation or pre-treatment of propagules	Poor plant establishment	Use trained personal and follow clear methodologies		Monitor all steps c measures to avoid to avoid interrupti or holidays). Prep (chemicals, tools)
	Failure in propagation and storage of propagules	Loss of germplasm			Group accessions procedures and v research for geno the general methc obtain additional i specific genotype
	Inadequate number of replicates per accession.	Loss of germplasm			Increase number represent the gen
<i>Field Bank Specifications</i>					
<i>Field Monitoring</i>	Unsuitable conditions in conservation site	Poor or suboptimal growth		Select a conservation site that is safe, favours plant development of the target germplasm, and isolated to prevent pest attacks and diseases but with easy access for management. Ensure that the climate and ecology of the site are conducive to maintenance.	
	High pest and disease pressure in field site	Loss of germplasm		Use screenhouse (SH) culture to provide the best protection against worst diseases, insects and pests.	
<i>Field Planting</i>	Pollen exchange with plants within and outside collection.	Loss of genetic integrity		Isolate site from potential pollinators if intended for outcrossing species.	Arrange the plant prevent plants fro reproductive struc pollinators, or use accession. Res rates of certain ta
	Misidentification	Loss of germplasm			Develop field map evaluation and ha name and access on maps. Use we computer-generat
	Mixtures of clones	Loss of genetic integrity			Provide adequate taking into consid of the plants. Pla rhizomes or runne between plots to p Accessions with c planted in adjacer spreading is a prc clones may requir to reduce mixing c accessions. Prun
	Contamination with volunteer plants.	Loss of genetic integrity			Use adequate cro grow after field pr before planting ne
<i>Field Maintenance & Management</i>	Mixtures of fruits and germplasms	Loss of genetic integrity			Conduct thinning overlapping betw and germplasms.

	Poor adaptation	Loss of germplasm			Monitor collection accessions to pos greenhouse or in to study and unde requirements of d better manage the
	Disparate location of physiologically similar accessions	Inefficient management			Plant accessions height, branching Crops that must b should be planted time to maturity.
	Poor management of weeds and low soil fertility	Loss of germplasm			Control weeds to weed-borne pathc fertility and adjust
<i>Post-harvest Handling</i>	Persistence of disease organisms and insects after harvest	Deterioration of propagules and spread of pests and diseases during storage			Treat tubers with storage. Closely r infections, and im prevent them from
	Mishandling	Deterioration of propagules during storage			Take extreme car transportation to e
Characterization and Evaluation	Inefficient and erroneous data gathering and encoding	Backlog and inaccurate characterization data	Assign staff with adequate training in characterization following international standards.	Provide digital hand-held encoder.	Independently ver computing, updati characterization d
	Descriptors that have no clear-cut correspondence to current international standard descriptors	No or limited usefulness of characterization data			Use updated desc all measurements
	Limited text-based description	Incomplete and inaccurate morphological description			Include images (6 accompanied with colors.
	Lack of diversity assessment of collection	Unknown level of breadth, duplication and gaps in collection, and conservation of unnecessary duplicates			Conduct molecu
DISTRIBUTION					
<u>Policies</u>	Lack of knowledge or negligence on germplasm exchange Protocol and International Treaty	Distribution without accompanying MTA. Inadvertent distribution of restricted germplasm (e.g. Non-MLS materials). Wrong information on the exchange status (MLS) of the germplasm.	Conduct regular update on international agreements concerning germplasm exchange.		Implement a clear distribution ensuri documents and aj are obtained befo
	Recipients of "designated" germplasm or "non-designated" germplasm attempt to claim IP rights over the germplasm	Restrictions on future access and use of germplasm			Distribute accessi CGIAR MTA for "i Center-created "n developed in colla CGIAR Center. w Center to take leg the MTA, upon re conditions.
	Plant health restrictions of importing country	Low level of exchange and utilization of germplasm.			Conduct research germplasm excha
	Non compliance with phytosanitary regulations	Germplasm distributed from genebank with diseases or pest contamination.			Test materials for compliance and g according to the p importing country. with an import per and a phytosanita
<u>User Service</u>	Germplasm distributed are weak	Dissatisfied recipients of germplasm			

Germplasm Preparation and Dispatch	Misclassification and wrong characterization and germplasm stocks data	Delayed identification and preparation of requested germplasm	Conduct regular training on germplasm characterization.		Check character identifiable charac relate to needs of Request all germ outside Center to related to the prov
	Inefficient and slow processing of requests for samples.	Dissatisfied recipients of germplasm	Dedicate personnel to serving germplasm requests.		Keep files of relev quarantine docum
	Errors in preparing or labeling samples	Wrong germplasm distributed by the genebank			Adopt barcoding & distribution protoc
	Insufficient germplasm stock for distribution	Delay in serving germplasm request			Incorporate alerts stock control syst multiplication and popular genetic st their DNA sample
	Bulky tissue-cultured explants	Expensive shipping cost and vulnerability of material to disintegration			Use space-saving up.
	Unfavorable conditions during transport	Delay in delivery , reduction of viability or loss of materials			Use packing mate unfavorable condi and under dry-ice shipment services
INFORMATION MANAGEMENT AND DISSEMINATION					
	Inefficient recording and database management	Backlog and inaccurate characterization data			Use GRU Databa to SINGER every in books.
	Mishandling of information and disorganized data sets (e.g. information system, field/ lab observation)	Loss or inaccessibility of information			Use GRU Databa data sheets. Integ distribution recor policy databases.
	Improper recording of moisture content, germplasm inventory, viability, storage location, and characterization data.	Inaccurate or wrong information			Independently ver computing, updati inventory, viability characterization d making tools in th genebanking ope
	Lack of adequate information about important characteristics of each accession.	Low interest and utilization of germplasm			Collect data on irr information from
	Mislabelling of new bags and other containers for the germplasm accession and samples are placed in the wrong container.	Loss or misplacement of materials			Set up a standard placement of sam regeneration to ha an explant throug mixture of letters ; possibility of trans barcoding system Use preprinted lab scanners and poc labeling that does the label. Maintair accession as a re
	Lack of secure back-up	Loss of genebank data		Transfer new data on CD or tape in two central databases kept in separate buildings in the institute. They can be stored also in secure, passport-regulated cyberspace.	Produce hard phc original data shee transaction comp daily incremental ups.
	Important data and information remain in useless form.	Low level of utilization of germplasm and information.			Disseminate relev germplasm and g in germplasm cat bulletins, and ope media.

Outdated or inaccessible procedures manual	Loss of improvements in procedures			Write out in detail procedures manual for all workers. Update the manual as needed. Update the major procedures manual.
Inconsistent protocols	Much variation in quality of process outputs			Develop standard media sheets as well as standard protocols. When new protocols are developed, file the old ones for reference.
Lack or complicated tracking and inventory system	Loss or misplaced samples and failure to regenerate and serve germplasm request on time			Design a computerized tracking and inventory system for researchers to follow. Acquisition through the system should include information, field collection records for each accession, location in dewar, meristems per via technique required, and important procedures.
Insufficient data on accession identity and culture conditions	Underestimate of germplasm viability or failure to propagate by recipient			Include each accession's initiation medium, growth medium, growth in length of subculture, and same numbering system as the genebank to allow the mother plant to be identified.
Limited ICT capability; server, network and IT related problems	Lack or poor accessibility of germplasm and important data to potential users	Engage a competent data curator to document decades of evaluation data in a centralized database system.	Use stable software and hardware and engage full technical support from Information Technology Unit. Change computers every 5 years. Upgrade memory and operating system every year.	Regulate software use. Restrict use of computers.
Malfunctioning equipment, hardware and software problems	Failure to update data by genebank staff. Delays in recording of accessions and declaring them to FAO & SINGER		Install redundant UPS units and hot-swappable battery packages. Enforce automatic start-up of generator within 30 seconds. Use alternating 2 power-supplies connected to the same server.	Enable immediate access to home phones in case of problems. Backup data kept in Center, then in the field and later in the internet.
INFRASTRUCTURE/PHYSICAL FACILITY				
Storage conditions at genebank not suitable (temperature, humidity, light conditions, exposure to contaminating organisms, pests)	Reduction or loss of viability		Treat culture rooms with pesticides on a regular time basis. Regularly check and maintain cooling units. Maintain storage room conditions and monitor conditions daily via remote sensing devices. Install a High Efficiency Particle-removal Air system (HEPA) and alarm systems for open doors, temperature/ humidity changes in the culture areas. Provide a dehumidifier.	Conduct regular monitoring for fungus problems. Monitor MTS materials for occurrence of necrosis, blackening, contamination, and defoliation. Develop a protocol for handling such problems.
Poor organization of storage trays, shelves and compartments	Loss or misplacement of germplasm	Restrict storage facility access to authorized genebank personnel.	Rationalize arrangement of storage trays, shelves, and compartments.	Develop a simple system for space units. Conduct regular verification of location of accessions on computerized database.
Deterioration of facilities and equipment	Reduction or loss of viability		Pursue continual upgrading and expansion of field and laboratory equipment, etc.	
Cold room malfunction	Reduction or loss of viability		Place hygrothermographs that are connected to back-up power supply and alarm system. Provide the rooms with multiple compressors and dehumidifiers that are programmed for alternate operation.	

	Power supply cut-off	Reduction or loss of viability		Install, regularly check, and maintain an emergency electrical generator for back-up power to the storage rooms, essential genebank lighting, monitoring devices, and access locks during electrical power failures.		
	Theft or vandalism	Loss of germplasm		Place the building under 24-hr perimeter security surveillance. Link the alarm system by optical fiber with security office and police. Install double locks in sensitive areas and closed-circuit camera monitoring by guards. Install sensors for door contacts, glass breaks and unusual motion outside work hours.	Restrict access to personnel with access. Conduct who will use facility on the safety and genebanks.	
	Environmental risks/weather elements, earthquakes, other catastrophic events (civil war,...), and fire	Reduction or loss of viability	Assign personnel from genebank unit and security office for 24/7 watch of the facility.	Design and construct building according to safety, environmental and artillery protection, and earthquake proof standards. Install automatic fire and gas alarm systems and provide fire isolation doors and fire extinguishers. Provide doors than can open from inside cold chambers to prevent personnel getting trapped.	Conduct periodic genebank during leaks in the cold a check fire safety	
<i>Safety Duplication</i>	Safety duplication site is vulnerable to natural calamities	Inaccessible or loss of safety duplication		Establish duplicate back-up in a geologically secure site with low radiation (radioactivity) and stable (low probability of earthquakes). The facility must be situated at an altitude that guarantees proper drainage during seasonal rains and eliminates the risk of flooding in the event of rising sea levels due to global warming.		
	Changing policies, financial and technical capabilities of governments hosting safety duplication	Inaccessible or loss of safety duplication		Establish safety backup arrangements in two different, economically stable countries, preferably in different continents, for black-box storage. Prepare a pull-out scheme in the event of instability in host country. Duplicate collection in two other sites, or keep an in vitro or a cryo set.		
PERSONNEL AND SUPPORT SERVICES						
	Inadequate complement of technical staff	Inefficient operations		Hire at least one highly qualified technician each to manage germplasm viability test, germplasm drying and moisture test, germplasm health test, characterization and regeneration, data management, and germplasm distribution. For an active collection with research and development needs, hire a scientist to take charge of planning, research and analysis, a technician to take charge of daily operation of the laboratory, laboratory assistants for germplasm cleaning, germplasm processing and germplasm packaging, and field workers for germplasming, field-layout, screenhouse and field maintenance and harvesting.		
	Incompetent staff	Inefficient operations		Hire researchers with advanced degrees in plant physiology/genetics. Hire laboratory technicians with a background in plant science. Hire laboratory assistants with training in basic botany. Provide 1-2 weeks intensive on-site training for each new staff member on standardized laboratory and field protocols, followed by close supervision for as long as needed.		

Routine tasks and uncompetitive remuneration	Fast staff turnover	Rotate work assignments as much as possible or assigning special projects to laboratory assistants. Train each assistant to make medium, wash dishes, transfer cultures, check cultures for contaminants, do basic record keeping, and other required laboratory tasks. Educate workers on the mission of the facility to provide a morale boost and establish a research-oriented approach to work.		
Exposure to occupational hazards	Reduced manpower capability		Provide protective clothing, gloves and safety devices such as showers, eyewash and fire extinguishers.	Protect staff members, for example, by spraying
Suffocation/asphyxiation and frostbite and cold injury from LN exposure. Mechanical injury incurred on explosion of a pressurized vessel containing LN.		LN safety considerations should be included in the training of all new staff.	Well-ventilated room; handling and storage dewars must be vented; skin and eyes must be protected with cold-resistant gloves, aprons, safety glasses and closed-top shoes. Only LN-resistant vessels and instruments guaranteed by the manufacturer should be exposed to its vapor and liquid phases. Install oxygen level sensors and self-contained breathing apparatus. Install door magnetic locks that automatically unlock during emergencies.	Enforce safety system. Constantly monitor.
Inefficient human resources services	Delayed hiring of required manpower			Review and streamline
Inefficient purchasing and repair services	Delayed delivery/repair of required supplies and equipment			Review purchasing process. Keep spare equipment in stock (available), as a risk batteries, lamps, f
High cost of genebank operations	Loss of donor and user support			Closely follow and promote The Global Crop I donors.